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# **Protein Kinase A-dependent Neuronal Nitric Oxide Synthase Activation Mediates the Enhancement of Baroreflex Response by Adrenomedullin in the Nucleus Tractus Solitarii of Rats**

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## **Abstract**

### **Background**

Adrenomedullin (ADM) exerts its biological functions through the receptor-mediated enzymatic mechanisms that involve protein kinase A (PKA), or neuronal nitric oxide synthase (nNOS). We previously demonstrated that the receptor-mediated cAMP/PKA pathway involves in ADM-enhanced baroreceptor reflex (BRR) response. It remains unclear whether ADM may enhance BRR response via activation of nNOS-dependent mechanism in the nucleus tractus solitarii (NTS).

### **Methods**

Intravenous injection of phenylephrine was administered to evoke the BRR before and at 10, 30, and 60 min after microinjection of the test agents into NTS of Sprague-Dawley rats. Western blotting analysis was used to measure the level and phosphorylation of proteins that involved in BRR-enhancing effects of ADM (0.2 pmol) in NTS. The colocalization of PKA and nNOS was examined by immunohistochemical staining and observed with a laser confocal microscope.

### **Results**

We found that ADM-induced enhancement of BRR response was blunted by microinjection of NPLA or Rp-8-Br-cGMP, a selective inhibitor of nNOS or protein kinase G (PKG) respectively, into NTS. Western blot analysis further revealed that ADM induced an increase in the protein level of PKG-I which could be attenuated by co-microinjection with the ADM receptor antagonist ADM<sub>22-52</sub> or NPLA. Moreover, we observed an increase in phosphorylation at Ser1416 of nNOS at 10, 30, and 60 min after intra-NTS administration of

ADM. As such, nNOS/PKG signaling may also account for the enhancing effect of ADM on BRR response. Interestingly, biochemical evidence further showed that ADM-induced increase of nNOS phosphorylation was prevented by co-microinjection with Rp-8-Br-cAMP, a PKA inhibitor. The possibility of PKA-dependent nNOS activation was substantiated by immunohistochemical demonstration of co-localization of PKA and nNOS in putative NTS neurons.

### **Conclusions**

The novel finding of this study is that the signal transduction cascade that underlies the enhancement of BRR response by ADM in NTS is composed sequentially of cAMP/PKA and nNOS/PKG pathways.

### **Background**

Adrenomedullin (ADM), a 52-amino acid peptide, was originally isolated from human pheochromocytoma and initially shown to have potent vasodilatory activity [1]. The physiologic and pharmacologic functions of ADM have been intensively investigated after its discovery (for review see [2]). ADM exerts multiple biological activities by acting on its specific receptors, composed of calcitonin receptor-like receptor (CRLR) and receptor activity modifying protein (RAMP)-2 or -3 [3]. The hypotensive effect of intravenously administered ADM has been attributed to activation of ADM receptors (ADMRs) located on blood vessels [1]. In addition to distribution in the cardiovascular system, ADM and ADMRs are also expressed in the central nervous system (CNS) and are particularly localized to the autonomic nuclei, including nucleus tractus solitarius (NTS), lateral parabrachial nucleus (LPBN), and rostral ventrolateral medulla (RVLM) [4-6]. These findings suggested a possible role for ADM in central regulation of cardiovascular functions.

Indeed, several studies demonstrated that microinjection of ADM into the CNS induces brain area-specific changes in arterial pressure and heart rate (HR) [7-8]. Other studies further indicated that central ADM also exhibits an area-specific regulation on the baroreceptor reflex (BRR) in anesthetized or conscious animals [9-11]. In our recent study [12], we demonstrated that microinjection of ADM into NTS, the termination site of primary baroreceptor afferents in the brain stem [13], significantly increases BRR response and sensitivity in a time- and dose-dependent manner, without producing discernible changes in basal arterial pressure and heart rate.

Stimulation of cyclic adenosine monophosphate (cAMP) formation is suggested to be the primary downstream mechanism subsequent to activation of the  $G_s$  protein-coupled ADMRs in vascular cells [1,14]. In CNS neurons, the cAMP-associated mechanism is also considered to be the primary signaling pathway that mediates ADM actions. Xu and Krukoff reported that ADM inhibits the baroreflex control of HR via activation of cAMP-dependent protein kinase A (PKA) in RVLM of the rat [11]. Our previous study also revealed the involvement of cAMP/PKA-dependent mechanism in BRR augmentation in response to activation of ADMRs in NTS [12]. In addition to cAMP/PKA pathway, nitric oxide (NO) has been suggested to serve as another intracellular signaling molecule that mediates the ADM actions [2]. In RVLM and LPBN, ADM induces hypertensive effect through cyclic guanosine monophosphate (cGMP)-associated signaling that is mediated by NO derived from neuronal NO synthase (nNOS) [15-16]. However, whether the nNOS-dependent mechanism contributes to the BRR-enhancing effect of ADM in NTS remains unclear.

The present study was undertaken to evaluate the hypothesis that ADM may enhance BRR through PKA-dependent activation of nNOS in NTS. Our results support this hypothesis and reveal that nNOS may mediate ADM-induced BRR enhancement via activation of cGMP-dependent protein kinase G (PKG) in NTS. We further found that a PKA-dependent phosphorylation at the amino acid residue Ser1416 accounts for the ADM-induced nNOS activation.

## **Materials and methods**

### **Animals**

Sprague-Dawley rats (male, weighing 300–400 g) obtained from the Animal Center of National Yang-Ming University were used in this study. Rats were housed in a laboratory animal room under controlled temperature (25°C) and light on 0800–2000 h, and had unrestricted access to food and water. All animals were allowed to acclimatize for at least 3 days before use. Animal care and all experimental protocols applied in the present study were approved by the Institutional Animal Care and Use Committee of National Yang-Ming University.

### **Surgical preparation**

As described previously [12], rats were anesthetized by intraperitoneal (i.p.) injection of pentobarbital sodium (50 mg/kg) and placed on a heating pad. The trachea was intubated to facilitate ventilation, and the femoral artery was cannulated for monitoring systemic arterial pressure (SAP). The femoral veins on both sides were also cannulated for injection of test agents and administration of supplemental anesthetics. Mean AP (MAP, mmHg) and HR (beats/min) were derived from the pulsatile SAP signals measured with a

pressure transducer (T844, ADInstruments, Castle Hill, Australia). To provide satisfactory anesthetic maintenance [17], rats received continuous infusion of pentobarbital at a rate of 15-20 mg/kg/h throughout the recording session.

### **Microinjection**

The rat was placed in a stereotaxic frame (Kopf, Tujunga, CA, USA) followed by an occipital craniotomy to expose the dorsal surface of the medulla. A glass pipette adapted to a Hamilton microsyringe (Reno, NV, USA) was used to microinject test agents into NTS. The coordinates used were: 0.5 mm rostral to the calamus scriptorius,  $\pm 0.5$  mm lateral to the midline, and 0.5 mm below the surface of the medulla. The volume of injection was limited to 20 nl per site. For histological verification of injection sites, the microinjection medium for test agents or artificial cerebrospinal fluid (aCSF) contains 1% Evans blue.

### **Test agents**

ADM was purchased from Bachem AG (Hauptstrasse, Bubendorf, Switzerland); N-propyl-L-arginine (NPLA), S-methylisothiourrea (SMT), L-NIO or 8-bromo-cAMP (8-Br-cAMP) from Tocris (Bristol, UK); Rp-8-bromo-cAMP (Rp-8-Br-cAMP) or Rp-8-bromo-cGMP (Rp-8-Br-cGMP) from Calbiochem (San Diego, CA, USA); ADM<sub>22-52</sub>, 3-morpholinolinosyndnomine (SIN-1), S-nitrosoglutathione (GSNO) or phenylephrine from Sigma-Aldrich (St. Louis, MO, USA); and L-NAME from Cayman (Ann Arbor, MI, USA).

### **Measurement of BRR response**

The procedures and methods for measuring the BRR response were described previously [12]. In brief, a bolus intravenous injection of phenylephrine (10  $\mu$ g/kg) was administered to evoke the BRR before and 10,

30 or 60 min after microinjection of the test agent into NTS. The BRR response was represented by the ratio of the peak magnitude of reflex bradycardia to the peak magnitude of phenylephrine-induced pressor response. The averaged value of BRR response obtained from three injections of phenylephrine prior to microinjection of the test agent served as the baseline control.

### **Histology**

At the end of the physiological experiments, animals were killed with a high dose of pentobarbital sodium (100 mg/kg, i.p.). The brain stem was then removed and fixed in 10% paraformaldehyde-saline solution that contains 30% sucrose for 48-72 h. Serial sections were cut (20  $\mu$ m) in a cryostat (Leica, Wetzlar, Germany) and mounted on slides. The sections were then stained with neutral red, and the microinjection site (marked with Evans blue) was identified under a microscope.

### **Immunofluorescence staining**

The procedures of triple immunofluorescence staining were described in a previous study [12]. Briefly, rats were deeply anesthetized and perfused transcardially with warm heparinized saline, followed by ice cold 4% paraformaldehyde (pH 7.4). Brains were then rapidly removed and postfixed at 4°C overnight. The medulla oblongata at the level of obex was sectioned coronally at a thickness of 10  $\mu$ m. Sections were then incubated with a mouse anti-nNOS antiserum (1:25; Santa Cruz Biotechnology, Santa Cruz, CA USA) and a rabbit anti-PKA antiserum (1:50; Santa Cruz Biotechnology) for 24 h at 4°C followed by 1-h incubation of Alexa Fluor 546-conjugated goat anti-mouse IgG (1:125; Invitrogen, CA, USA) and Alexa Fluor 488-conjugated

donkey anti-rabbit IgG (1:250; Invitrogen). Nuclear staining was performed with 4'-6-diamidino-2-phenylindole (DAPI) (1:250; Invitrogen, Carlsbad, CA, USA) in PBS for 10 min at room temperature. Immunoreactive expression of proteins was observed with a laser confocal microscope (Leica, Wetzlar, Germany).

### **Western blotting**

The experimental protocols for Western blot analysis of ADM-induced protein expression were described previously [12]. In brief, tissues from separate groups of rats obtained 10, 30 or 60 min after bilateral microinjections of aCSF or test agents into NTS were collected. The tissues covering the anatomical boundaries of the dorsomedial NTS were visualized and micropunched with the aid of a dissecting microscope. After tissue homogenization and protein quantification, proteins of interest were separated using a 12% SDS-polyacrylamide gel and transferred to a polyvinylidene difluoride (PVDF) membrane. Following blocking of non-specific binding, membranes were incubated with a rabbit anti-PKG-1 $\alpha$  antiserum (1:2000; Calbiochem/EMD Biosciences, Darmstadt, Germany), a rabbit anti-nNOS antiserum (1:1000; Santa Cruz Biotechnology), a rabbit anti-phospho-nNOS (Ser847) antiserum (1:3000; Abcam, Cambridge, UK), a rabbit anti-phospho-nNOS (Ser1416) antiserum (1:3000; Abcam), or a rabbit anti- $\beta$ -actin antiserum (1:10000; Santa Cruz Biotechnology) in Tris buffer at 4°C overnight. This was followed by incubation with horseradish peroxidase (HRP)-conjugated donkey anti-rabbit IgG (1:10000; Santa Cruz Biotechnology) for 1 h at room temperature. Western blots were quantified by

densitometer and the relative density of proteins of interest was normalized against  $\beta$ -actin.

### **Statistical analysis**

All data are presented as mean  $\pm$  S.E.M. Results were analyzed by one-way or two-way ANOVA with repeated measures for group means, as appropriate, followed by Scheffe's post hoc test for individual means.  $P < 0.05$  was taken as statistically significant.

## **Results**

### **Involvement of nNOS in BRR-enhancing effect of ADM in NTS**

In our previous study [12], we have demonstrated that microinjection of ADM (0.2 pmol) into NTS significantly augmented BRR response with a maximal enhancement at 60 min after administration. Our first set of experiments established the participation of nNOS in this process. Microinjection bilaterally of ADM (0.2 pmol) into NTS elicited a 1.4 fold increase in the BRR response (Fig. 1A). L-NAME (25 pmol) blunted completely the BRR-augmenting effect of ADM (0.2 pmol) when co-microinjected with ADM (Fig. 1A). However, L-NAME, when given alone to NTS at 25 pmol, exerted minimal effect on BRR response (Fig. 1A). Comparable effects were obtained on co-microinjection of ADM with NPLA (250 pmol), a selective nNOS inhibitor (Fig. 1B). On the other hand, the ADM-induced BRR enhancement was substantially unaffected by co-microinjections with the relatively selective inducible NOS (iNOS) inhibitor SMT (250 pmol) (Fig. 1C), or L-NIO (100 pmol), a preferential endothelial NOS (eNOS) inhibitor (Fig. 1D).

### **nNOS-dependent PKG activation by ADM and in NTS**

Since PKG can be activated by nNOS-derived NO [18], we next examined the role of PKG in the BRR enhancement response induced by ADM in NTS.

Figure 2A shows that co-microinjection of Rp-8-Br-cGMP (1 nmol), a selective PKG inhibitor, abolished the ADM-elicited BRR augmentation. Western blot analysis revealed that ADM significantly increased PKG-I level in NTS 30 min after application, and was diminished by the ADMR antagonist ADM<sub>22-52</sub> or NPLA (Fig. 2B).

### **Phosphorylation of nNOS by ADM in NTS**

Phosphorylation at critical amino acid residues is important for the regulation of nNOS activity [19]. Since ADM induces dephosphorylation of nNOS at Ser847 and stimulates NO production from cultured hypothalamic neurons [20], we examined the effect of ADM on phosphorylation of nNOS at Ser847. As shown in Fig. 3A, the protein level of total nNOS was not substantially changed 10, 30, and 60 min after ADM administration. We also found that the protein levels of phospho-nNOS (Ser847) were not significantly altered during the time-period when BRR response was augmented by ADM (Fig. 3A & B).

In NTS, insulin-mediated cardiovascular effect was reported to involve the nNOS activation via phosphorylation at Ser1416 [21]. As illustrated in Fig. 3A, ADM induced a significant increase in protein levels of phospho-nNOS (Ser1416) in NTS. The ADM-induced increase in phosphorylation of nNOS at Ser1416 maximized at 30 min and gradually declined within 60 min after ADM administration (Fig. 3B).

### **PKA-dependent activation of nNOS induced by ADM in NTS**

In addition to the nNOS/PKG pathway, we demonstrated previously that the cAMP/PKA mechanism mediates the effects of ADM on baroreflex in NTS in

rats [12]. Since both PKA [12] and nNOS (Fig. 1) inhibitors abolished completely the ADM-elicited augmentation of BRR response, it is plausible that an in-series relationship exists between PKA and nNOS signaling pathways in the mediation of ADM effects in NTS. Our fourth series of experiments was carried out to examine whether nNOS phosphorylation is dependent on PKA activation evoked by ADM in NTS. We found that ADM-induced increase in phospho-nNOS (Ser1416) level was completely suppressed by co-microinjection with the PKA inhibitor Rp-8-Br-cAMP into NTS, while the level of total nNOS remained unaltered (Fig. 4).

We then verified the contribution of PKA-mediated nNOS activation to BRR augmentation. As illustrated in Fig. 5A, both 8-Br-cAMP (400 pmol), a PKA activator, and SIN-1 (1 nmol), a putative NO donor, mimicked the BRR-enhancing effect of 0.2 pmol ADM at 10-60 min after microinjection into NTS. We further found that the BRR enhancement induced by 8-Br-cAMP was completely blocked by L-NAME (Fig. 5B). On the other hand, the BRR-enhancing effect of SIN-1 was not altered by the PKA inhibitor Rp-8-Br-cAMP (Fig. 5B). Of note is that the BRR augmentation by microinjection of GSNO (0.5 nmol), a specific NO donor, was comparable to that of SIN-1 and was also unaffected by co-microinjection with Rp-8-Br-cAMP (Fig. 5C).

To determine whether nNOS and PKA are co-localized at the same NTS neuron, double immunohistochemical staining for nNOS and PKA proteins was carried out in rat brain slices. As shown in Fig. 6, putative NTS neurons positively expressed nNOS-immunoreactivity (IR) were also stained with immunofluorescence for PKA protein, while some neurons manifested PKA-IR alone.

## Discussion

The present study unveiled two novel findings. We found that the activation of nNOS/PKG cascade is responsible for BRR enhancement induced by microinjection of ADM into the NTS. We further showed that activation of nNOS by ADM is via a PKA-dependent mechanism. Together with our previous findings [12], this study demonstrated that the signal transduction cascade that underlies the enhancement of BRR response by ADM in NTS is composed sequentially of cAMP/PKA and nNOS/PKG pathways (Fig. 7).

This is the first report that provides direct biochemical and pharmacologic evidence to show that PKG-I, the active form of PKG, in NTS was upregulated by ADM-induced nNOS activation. NO participates in a wide variety of neuronal functions in the CNS, including cardiovascular regulation, nociception, synaptic plasticity, and control of complex behavioral responses (for review see [22]). At the NTS level, NO has been suggested to affect neuronal discharge and modulate the BRR response of the rat [23-26]. Although all three NOS isoforms have been suggested to be present in the NTS [27-28], the possibility that the activation of iNOS and eNOS may be involved in ADM-induced BRR-enhancing effect is deemed unlikely (Fig 1C & 1D). Moreover, several lines of evidence support the notion that nNOS-derived NO in NTS plays important physiologic roles in regulating transmission of arterial baroreflex signals and cardiovascular functions [29-30]. We further demonstrated in this study that the BRR-enhancing effect of ADM is mediated by nNOS-dependent PKG activation in NTS. We noted that the ADM-induced increase in nNOS phosphorylation declined gradually at 60 min after ADM microinjection, while the BRR-enhancing effect was sustained at the comparable time period. The discrepancy of temporal changes in

nNOS activity and BRR response may reflect the sequential participation of nNOS and its downstream molecules including PKG in the ADM-activated signaling cascades. The significance of nNOS-dependent PKG activation in BRR regulation is further substantiated by a previous study [18] that revealed a significant nNOS-dependent upregulation of PKG-I protein in NTS following baroreceptor activation.

Another novel finding of the present study is that ADM may induce a PKA-dependent nNOS activation in NTS to enhance the BRR response. Both cAMP/PKA and NO/PKG mechanisms contribute to cardiovascular regulations by ADM in RVLM [11,16]. We further demonstrated these two signaling pathways exist in in-series in NTS. Our immunohistochemical results also showed that the PKA- and nNOS-dependent mechanisms could be activated in the same NTS neuron. We recognized that some PKA-labeled NTS neurons did not expressed nNOS signal (Fig. 6D). Recently, the extracellular signal-regulated kinase (ERK)-dependent signaling pathway in the NTS has been demonstrated to modulate cardiovascular functions [31]. It is thus possible that the PKA-dependent ERK signaling, which could be found in adipocytes activated by ADM [32], may serve as the downstream mechanism responsible for ADM-induced PKA activation in those NTS neurons expressing PKA-immunoreactivity only. The nNOS-IR is localized in neurons other than in glial cells [27] and is highly co-localized with soluble guanylate cyclase in NTS [33]. These results further substantiate our observations that ADM-activated nNOS/NO-cGMP/PKG cascades could be resided in the same NTS neuron.

The enzyme activity of nNOS has been demonstrated to be intimately associated with the state of phosphorylation at the amino acid residues Ser847 and Ser1416 [19]. For instance, phosphorylation of nNOS at Ser847 by calmodulin-dependent kinases results in a decrease of its enzyme activity [34]. On the other hand, protein phosphatase 2A-mediated dephosphorylation at Ser847 can lead to the activation of nNOS [35]. Recently, Xu and Krukoff demonstrated in an *in vitro* study that ADM significantly stimulated NO production from primary rat hypothalamic neurons by dephosphorylation of nNOS at Ser847 through a mechanism of PKA-dependent activation of phosphatases [20]. However, our results demonstrated that ADM induced an increase in phosphorylation of nNOS at Ser1416 but not at Ser847 in NTS neurons. The time course of nNOS phosphorylation is also compatible with the BRR-enhancing response induced by ADM. Since phosphorylation of nNOS at Ser1416, a known phosphorylation site for Akt (protein kinase B), is an alternative way to increase its enzyme activity [21], it is possible that Akt signaling may be involved in PKA-dependent nNOS phosphorylation and contributed to the ADM-induced BRR enhancement in NTS. This possibility, however, is subjected to further delineation.

## **Conclusions**

We have previously demonstrated an important role for ADM in BRR enhancement that is mediated by a PKA-dependent mechanism in the NTS [12]. In the present study, the effect of ADM on baroreflex was further suggested to involve the activation of nNOS in NTS. We conclude that the signal transduction cascade that underlies the enhancement of BRR response by ADM in NTS is composed sequentially of cAMP/PKA and nNOS/PKG

pathways. These findings may provide a new insight for our understanding of ADM-elicited signaling mechanisms and their cross-talk in central regulation of cardiovascular functions.

## **Competing interests**

The authors declare that they have no competing interests.

## **Authors' contributions**

DHTY and LCC participated in the design of this study and helped to draft the manuscript. YCS carried out the immunohistochemical experiments. YCC and ICH carried out the neurophysiologic and neuropharmacologic studies, and performed the Western blotting analysis. YJL and ICC participated in the interpretation of data and performed the statistical analysis. JCY conceived of the study, designed and coordinated the experiments, and drafted the manuscript. All authors read and approved the final manuscript.

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## Figure legends

### **Figure 1 Involvement of nNOS-dependent mechanism in the effect of ADM on BRR response.**

A: Temporal changes in BRR response of the rat that received bilateral microinjections into NTS of aCSF, L-NAME (25 pmol), ADM (0.2 pmol) or ADM plus L-NAME. B: Temporal changes in BRR response of the rat that received bilateral microinjections into NTS of aCSF, NPLA (250 pmol), ADM (0.2 pmol) or ADM plus NPLA. C: Temporal changes in BRR response of the rat that received bilateral microinjections into NTS of aCSF, SMT (250 pmol), ADM (0.2 pmol) or ADM plus SMT. D: Temporal changes in BRR response of the rat that received bilateral microinjections into NTS of aCSF, L-NIO (100 pmol), ADM (0.2 pmol) or ADM plus L-NIO. Data are presented as means  $\pm$  SEM, n = 6 to 8 animals per group. \* p<0.05 compared with control; † p<0.05 compared with the ADM group at 10, 30 or 60 mins.

### **Figure 2 Involvement of cGMP-dependent mechanism in the effect of ADM on BRR response.**

A: Temporal changes in BRR response of the rat that received bilateral microinjections into NTS of aCSF, Rp-8-Br-cGMP (1 nmol), ADM (0.2 pmol) or ADM plus Rp-8-Br-cAMP. Data are presented as means  $\pm$  SEM; \*p<0.05 compared with control; † p<0.05 compared with the ADM group at 10, 30 or 60 mins. B: Representative gels (inset) and quantified data showing changes in the protein level of active form PKG-I in NTS of the rat at 30 min after receiving bilateral microinjections into NTS of aCSF, ADM (0.2 pmol), ADM plus ADM<sub>22-52</sub> (0.2 pmol), or ADM plus NPLA (250 pmol). Quantified data are

presented as means  $\pm$  SEM. The mean value of sham-operated control rats is represented as Basal. \*  $p < 0.05$  compared with aCSF.

**Figure 3 ADM-induced increase in phosphorylation of nNOS.**

A: Representative Western blotting gels showing temporal changes in phosphorylation at Ser847 (p-nNOS<sup>S847</sup>) and Ser1416 of nNOS (p-nNOS<sup>S1416</sup>) at 10, 30, and 60 min after intra-NTS microinjection of ADM (0.2 pmol). B: Quantified data showing temporal changes in the protein level of phospho-nNOS in NTS of the rat that received bilateral microinjections into NTS of aCSF (Control; sampled at 10 min after aCSF administration), or ADM (0.2 pmol). Data are presented as means  $\pm$  SEM. \*  $p < 0.05$  compared with control.

**Figure 4 ADM-induced PKA-dependent phosphorylation of nNOS.**

Representative Western blotting gels showing changes in phosphorylation at Ser1416 of nNOS (p-nNOS<sup>S1416</sup>) at 10 min after intra-NTS microinjection of aCSF (control), PKA inhibitor Rp-8-Br-cAMP (PKAi; 1 nmol), ADM (0.2 pmol), or ADM plus PKAi. B: Quantified data showing changes in the protein level of phospho-nNOS in NTS of the rat that received bilateral microinjections into NTS of aCSF, PKAi (1 nmol), ADM (0.2 pmol), or ADM plus PKAi. Data are presented as means  $\pm$  SEM. \*  $p < 0.05$  compared with control; #  $p < 0.05$  compared with the ADM group.

**Figure 5 Involvement of PKA-dependent NOS activation in the effect of ADM on BRR response.**

A: Temporal changes in BRR response of the rat that received bilateral microinjections into NTS of ADM (0.2 pmol), 8-Br-cAMP (400 pmol), or SIN-1 (1 nmol). B: Temporal changes in BRR response of the rat that received bilateral microinjections into NTS of aCSF, ADM (0.2 pmol), 8-Br-cAMP (400

pmol) plus L-NAME (25 pmol), or SIN-1 (1 nmol) plus Rp-8-Br-cAMP (1 nmol).

C: Temporal changes in BRR response of the rat that received bilateral microinjections into NTS of aCSF, ADM (0.2 pmol), GSNO (0.5 nmol), or GSNO plus Rp-8-Br-cAMP (1 nmol). Data are presented as means  $\pm$  SEM, n = 6 to 8 animals per group. \*  $p < 0.05$  compared with control at 10, 30 or 60 mins.

**Figure 6 Co-localization of PKA and nNOS proteins in NTS.**

Confocal microscopic images of NTS showing immunofluorescence staining for nNOS (A, Alexa Fluor 546), PKA (B, Alexa Fluor 488), or cell nuclei (C, DAPI). The merged image (D) showing single staining for PKA (arrowhead) or double immunofluorescence staining (yellow color) for PKA and nNOS (arrows). Scale bar: 25  $\mu$ m.

**Figure 7 Schematic model of cellular mechanisms underlying the enhancement of BRR response by ADM in NTS.**

AC: adenylate cyclase; ADM: adrenomedullin; ADMR: adrenomedullin receptor; GC: guanylate cyclase; Gs: stimulatory GTP-binding protein; nNOS: neuronal nitric oxide synthase; PKA: protein kinase A; PKG: protein kinase G.

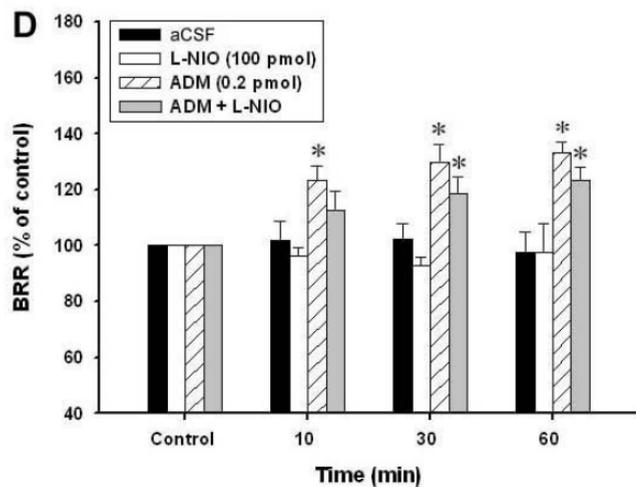
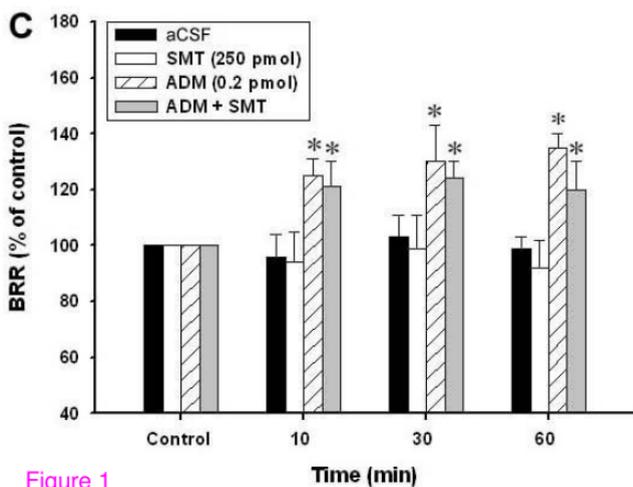
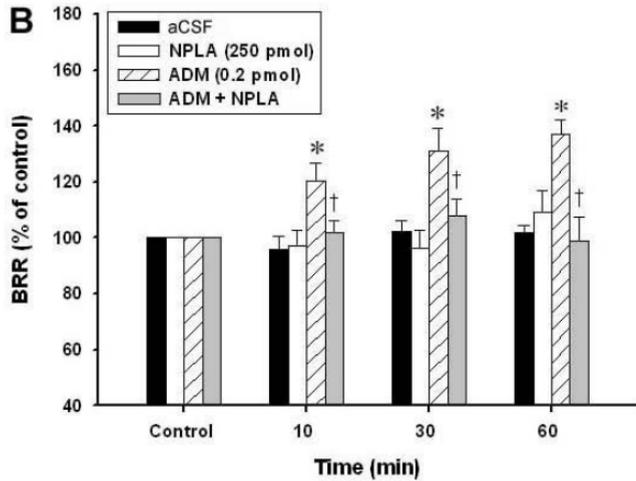
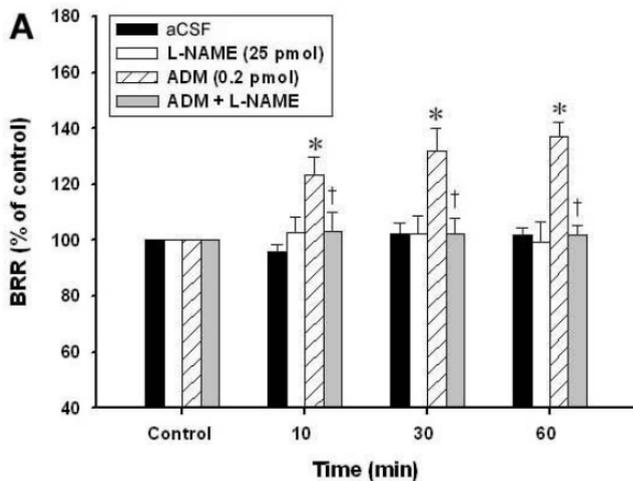


Figure 1

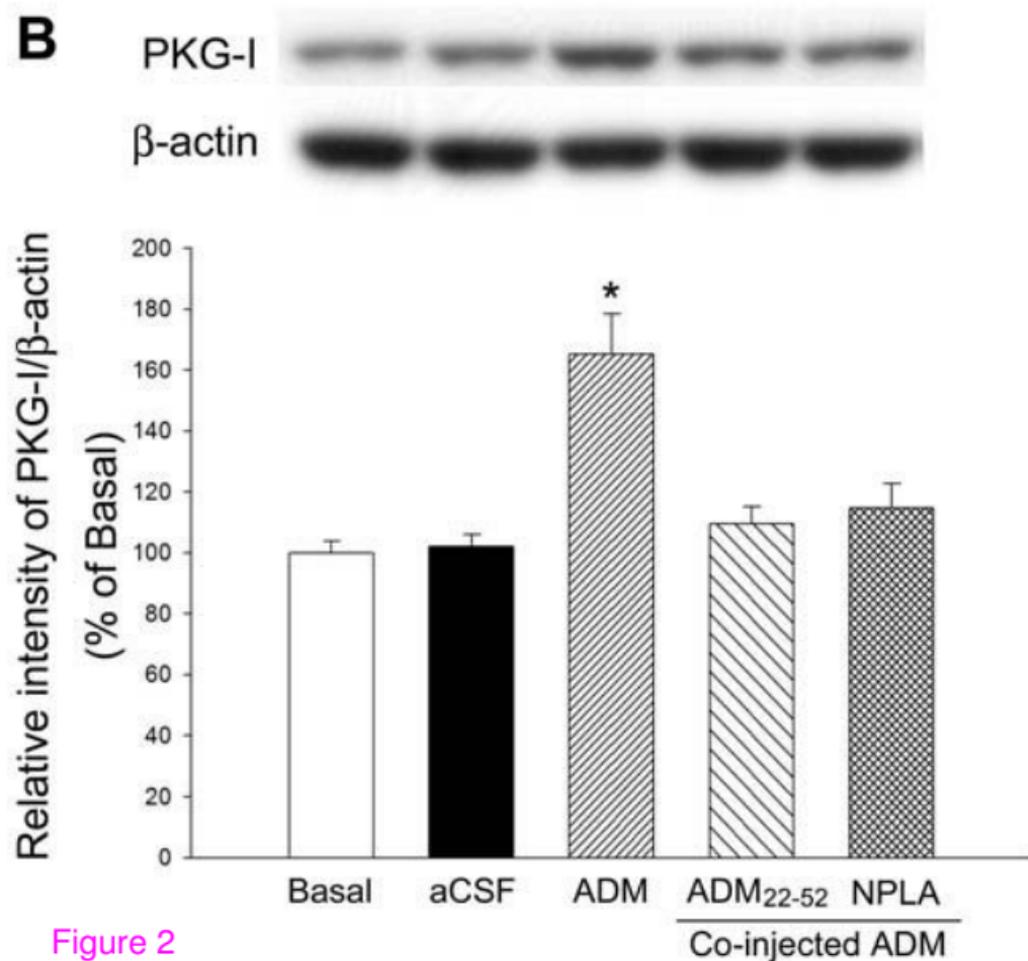
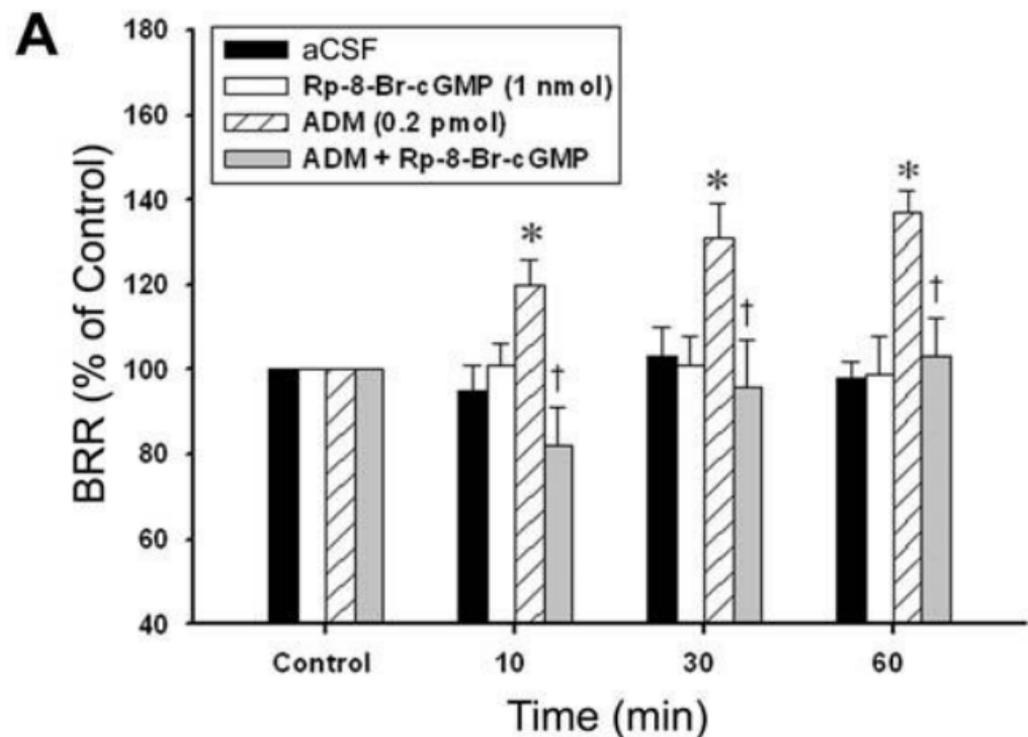


Figure 2

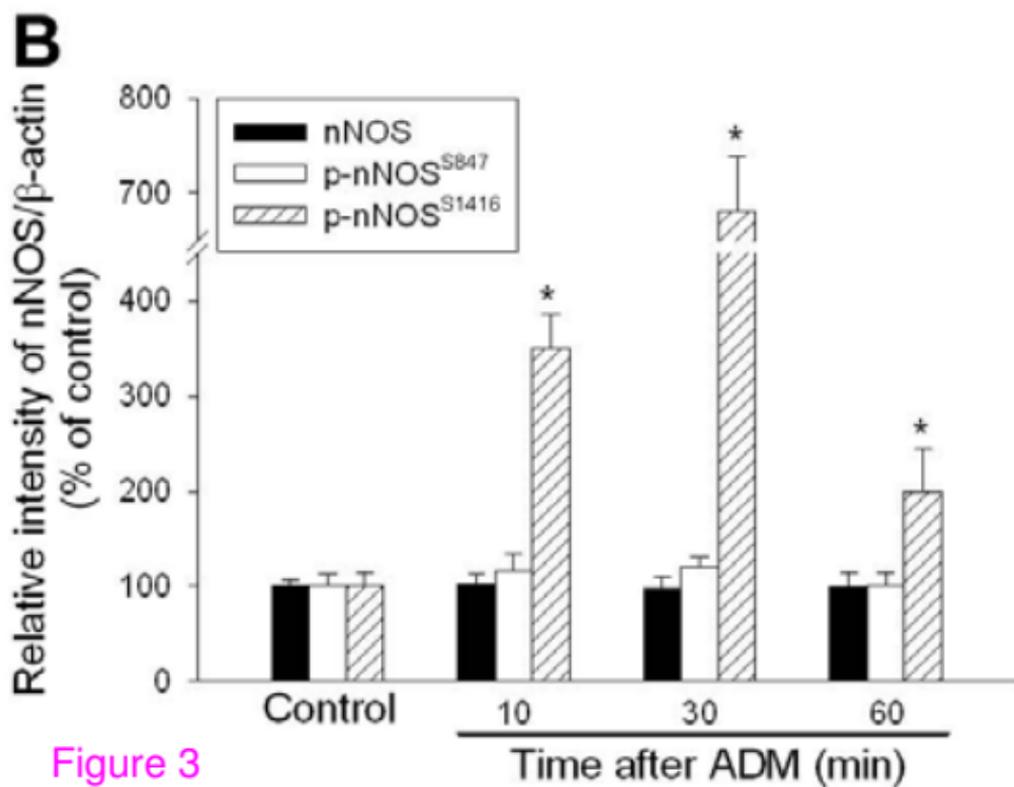
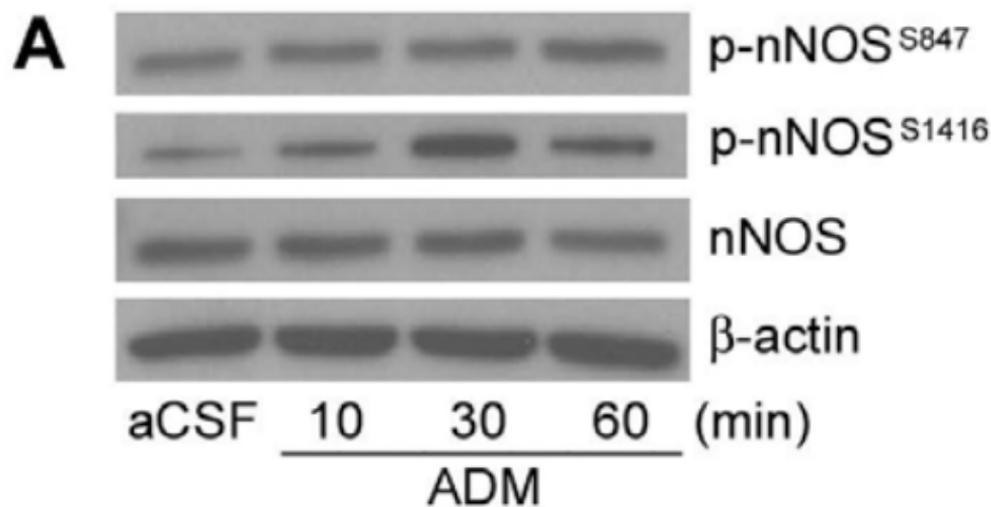


Figure 3

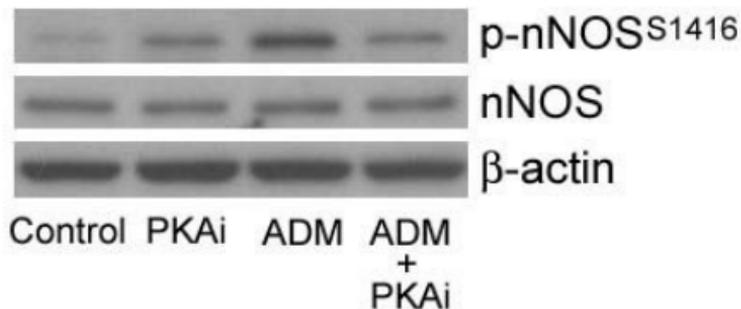
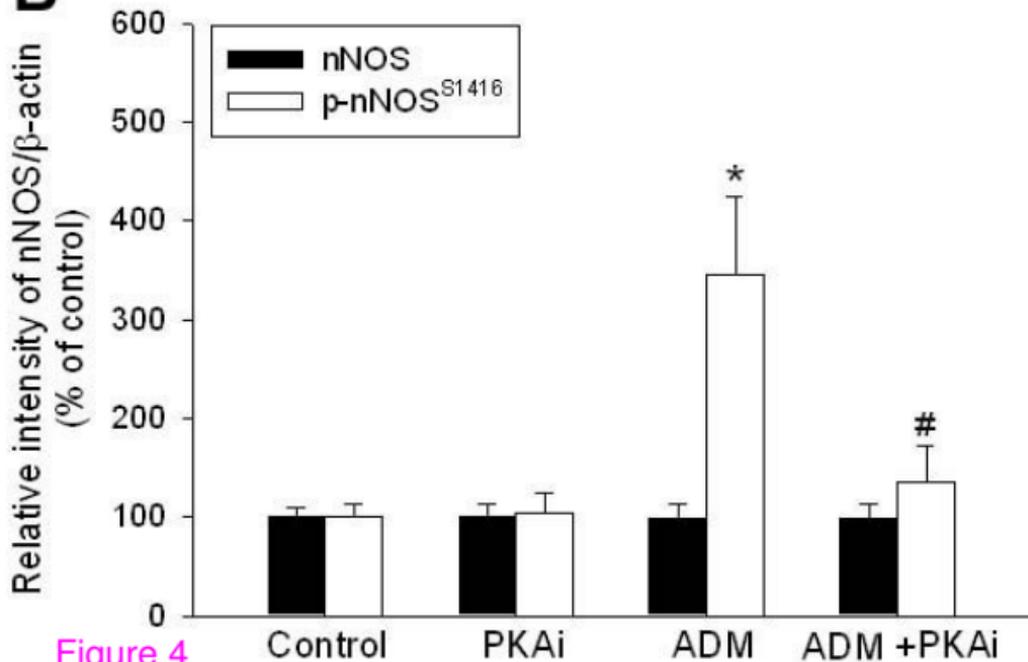
**A****B**

Figure 4

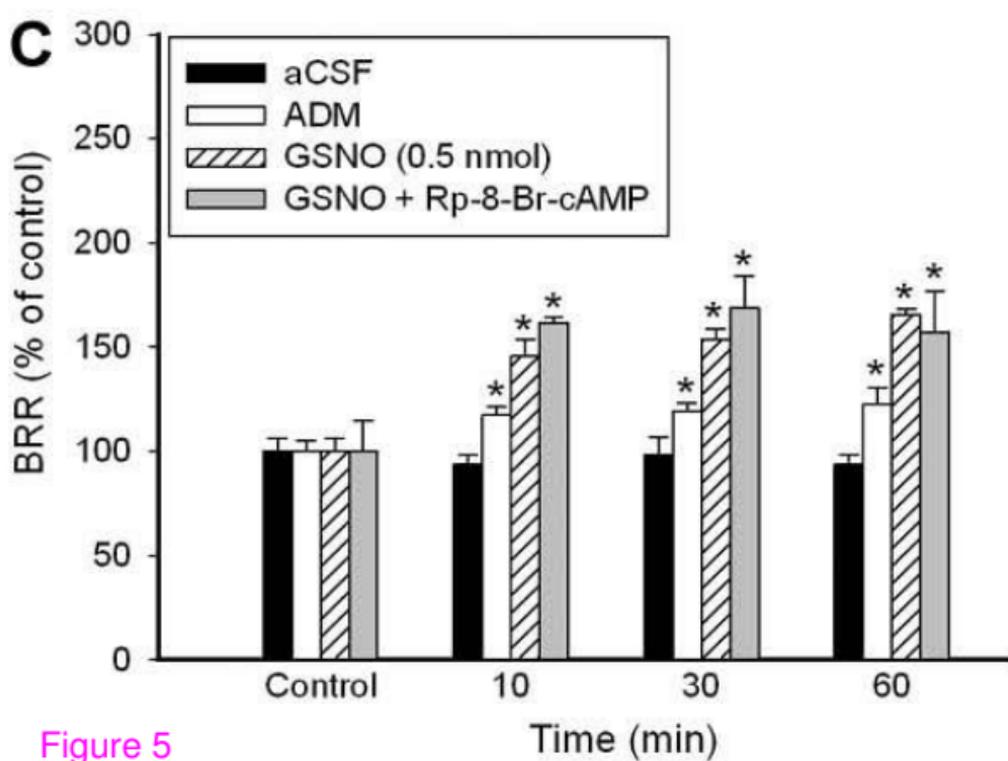
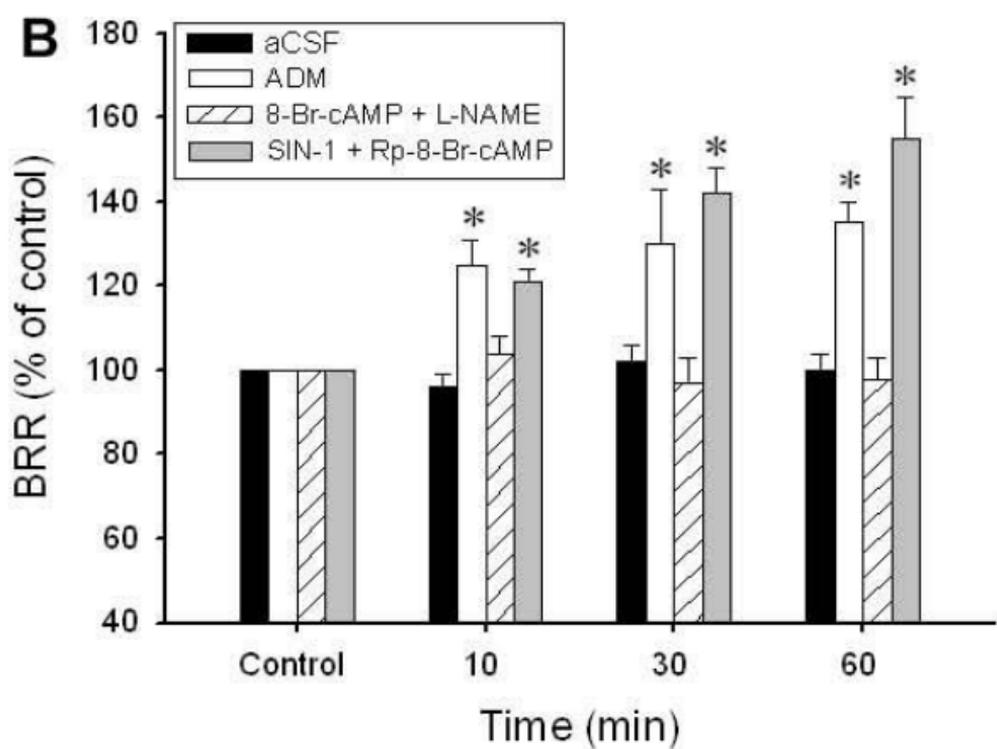
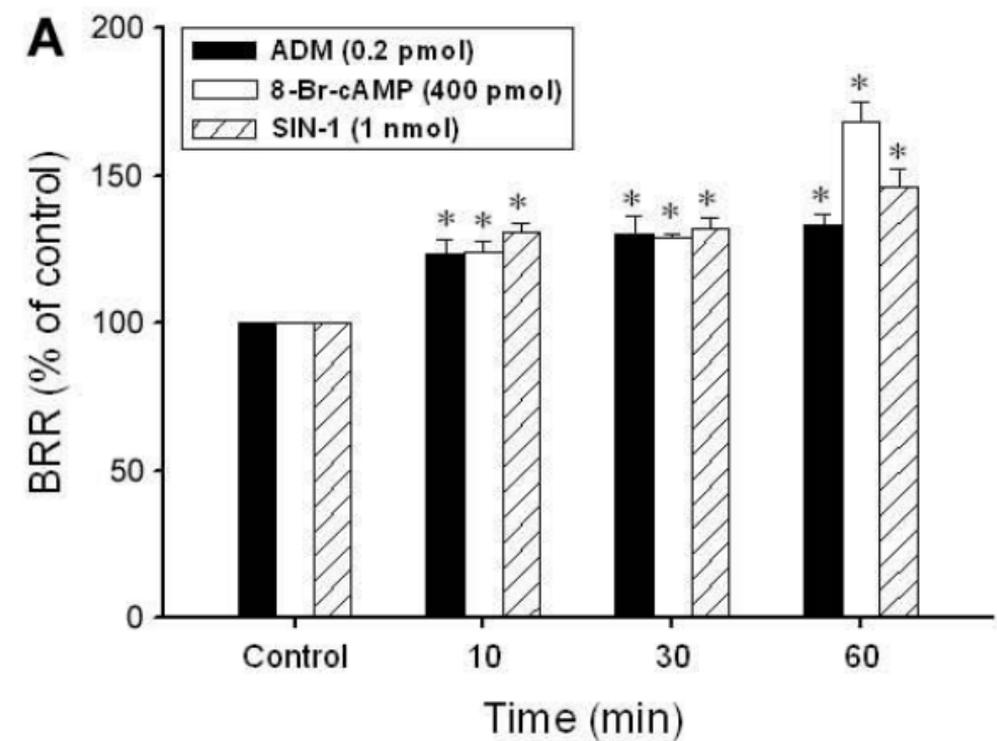


Figure 5

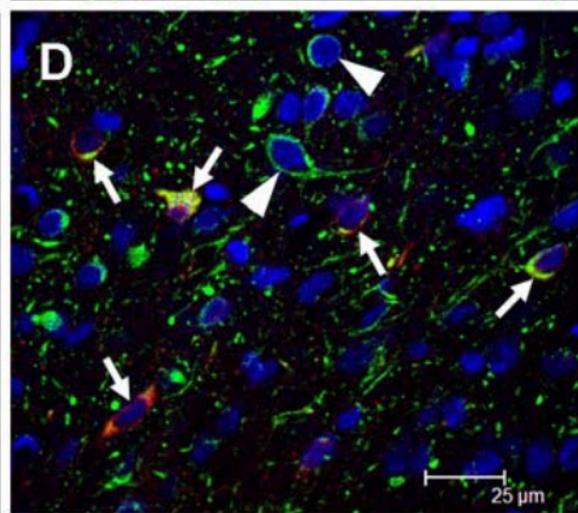
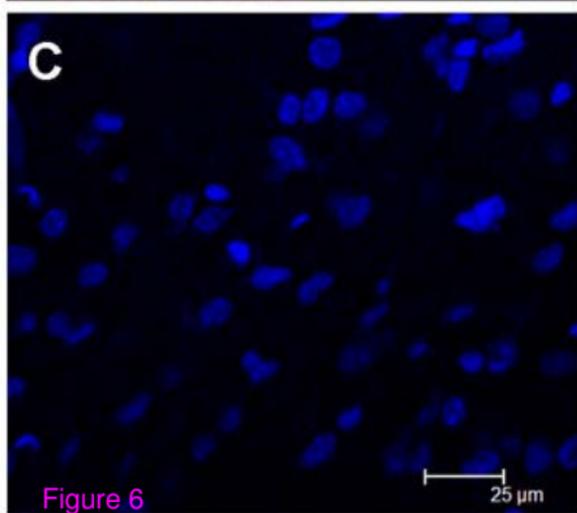
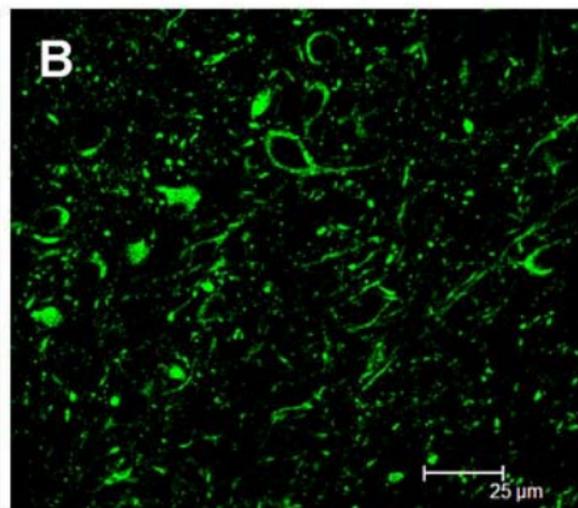
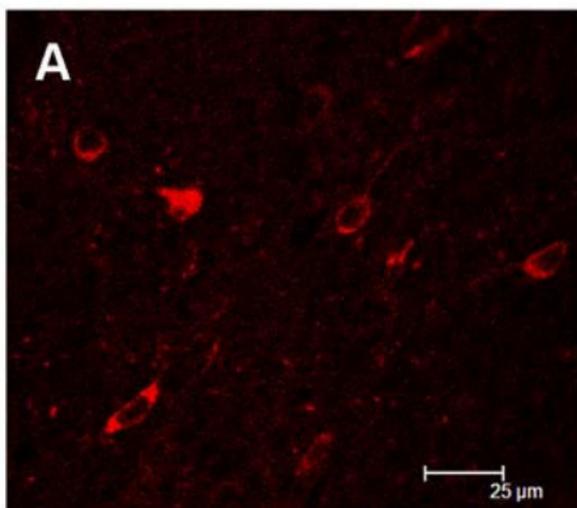


Figure 6

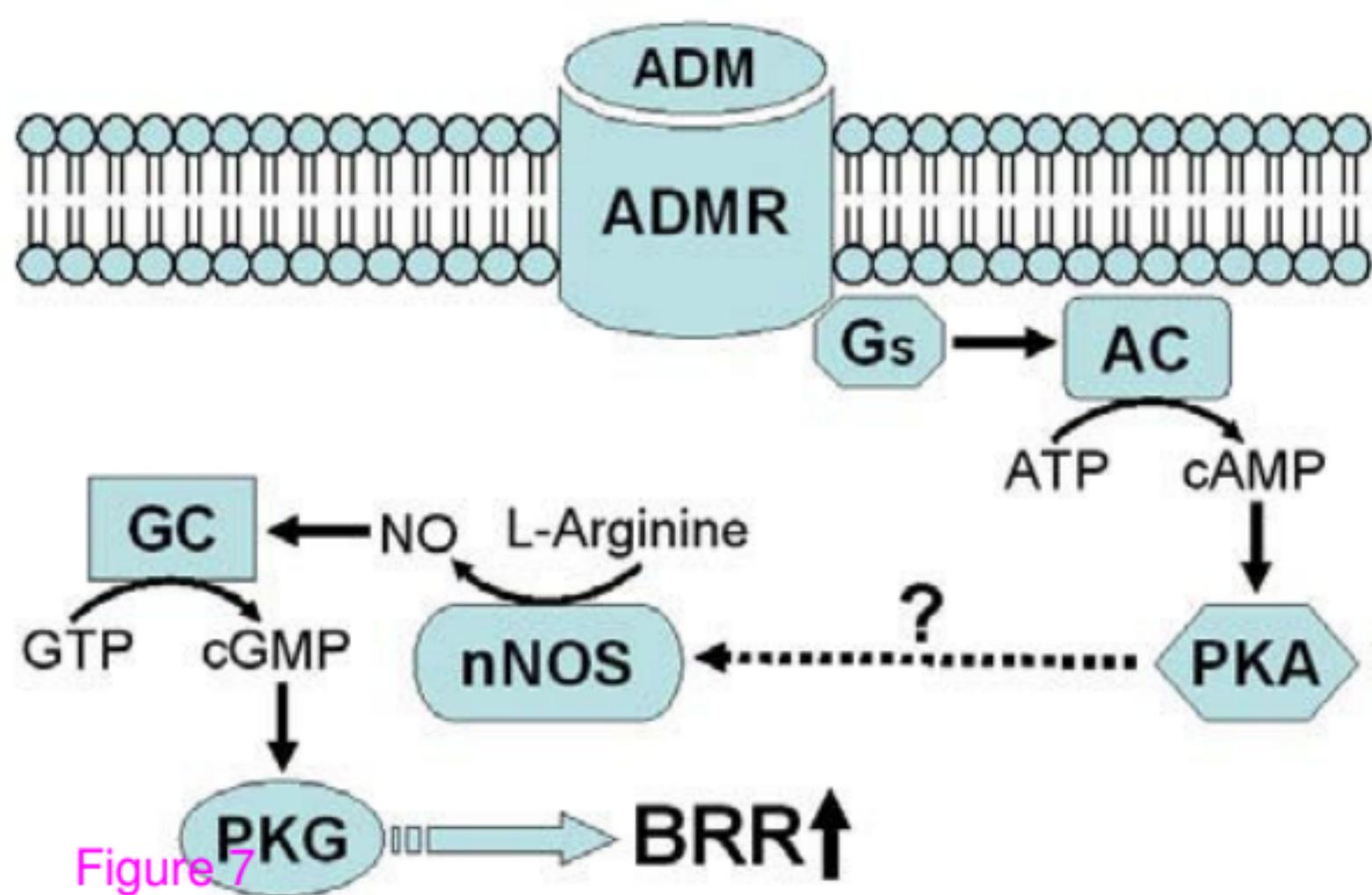


Figure 7